Enantioselective Synthesis of α -Silylamines by Meerwein–Ponndorf– Verley-Type Reduction of α -Silylimines by a Chiral Lithium Amide

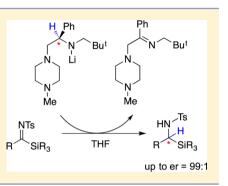
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Supporting Information

ABSTRACT: Meerwein–Ponndorf–Verley-type reduction of *N*-tosylsilylimines with chiral lithium amide **2** affords α -silylamines in high enantioselectivity. Since the enantioselectivity observed was inconsistent with our previously proposed chairlike sixmembered transition structure, we performed density functional theory (DFT) calculations on transition states leading to (*S*)- and (*R*)-7a and (*S*)- and (*R*)-7e using an *N*-phenylsulfonyl derivatives **12** and **13** as model systems. Results of the calculations showed that the structures are considerably deformed from the chairlike form with steric repulsions between the 1'-methylene group and the imine-carbon substituents playing an important role in the control of the enantioselectivity.



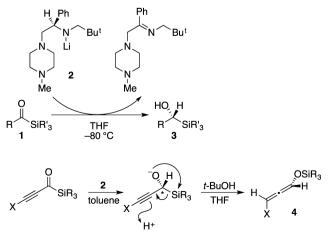
■ INTRODUCTION

Introducing silicon into bioactive compounds has attracted considerable interest in medicinal chemistry in view of potentially induced changes in conformations, lipophilicity, bonding property, and electronic nature.¹ One of the major applications for silicon-containing molecules is their use as a carbon isostere, in which, for example, silicon-based amino acids can act as an active tetrahedral intermediate of a protease inhibitor.^{2,3} From that of point of view, enantioselective synthesis of α -silylamines has recently attracted considerable attention.⁴ Although there have been many reports⁵ on the synthesis of racemic α -silylamines since the first preparation by Speier and Sommer in 1951,⁶ only a few methods for enantioselective synthesis of these compounds are available. The first report of an enantioselective synthesis of α -silylamines was by Tacke and Hengelsberg,⁷ who used enzymatic hydrolysis of *rac-N*-acyl- α -silylamines, derived from alkylation followed by amination of dimethylphenylsilylmethyl chloride. The reaction of enantioenriched α_{β} -epoxysilanes with sodium azide followed by LiAlH₄ was used for the preparation of α silylamines bearing a β -hydroxy group by Bassindale and coworkers.⁸ Picard and Fortis also reported the synthesis of an α silvlamine by reduction of the corresponding α -silvliminium chloride using lithium borohydride and diethyl tartrate.9 Although this method is applicable to aliphatic nonconjugated silylamines, the yield (43%) and enantioselectivity (60% ee) are moderate. A retro aza-Brook rearrangement (N to C) in Bocprotected N-silyl allyl, benzyl, or propargylamines in the presence of (-)-sparteine provides the corresponding α silylamines in >90% ee as reported by Sieburth.¹⁰ Scheidt and co-workers reported the synthesis of α -silylamines by the addition of silyl anions to chiral tert-butanesulfinimines.¹¹ Although this method was originally limited to non-enolizable

imines as substrates, Skrydstrup found some sulfinimine substrates with α -protons to be tolerated under the addition conditions.¹²

Fifteen years ago, we reported an enantioselective Meerwein–Ponndorf–Verley (MPV)-type reduction of acylsilanes¹³ by chiral lithium amide 2,¹⁴ in which hydride transfer occurs enantioselectively from a stereogenic carbon atom adjacent to a nitrogen atom of a chiral lithium amide to a carbonyl carbon atom $(1 \rightarrow 3)$ (Scheme 1). The method was recently applied to the enantioselective synthesis of silox-

Scheme 1. Enantioselective Reduction of Acylsilane Using Chiral Lithium Amide



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Tab	le 1.	Synthe	ses of	N-T	'osyl	limine
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		O H SiR₃ 5a-I	$\frac{\text{Et}_{3}\text{N} (4.4 \text{ equiv.})}{\text{CH}_{2}\text{Cl}_{2} 0 \text{ °C to rt}} R^{2}$	NTs ⊥ SiR₃ a-I	
entry	6	R	SiR ₃	time	yield $(\%)^a$
1	6a	Ph	SiMe ₃	1 h	67
2	6b	Ph	SiMe ₂ Bu ^t	2 h then reflux 2.5 h	74
3	6c	Ph	SiMe ₂ Ph	7 h	57
4	6d	Ph	SiMePh ₂	38 h then reflux 3 h	56
5	6e	TMSC=C-	$SiMe_2Bu^t$	reflux 7 h	97
6	6f	PhC≡C-	SiMe ₂ Bu ^t	14 h then reflux 8 h	83
7	6g	$Ph(CH_2)_3C \equiv C$ -	SiMe ₂ Bu ^t	reflux 8 h ^b	55
8	6h	TMSCH=CH-	SiMe ₂ Bu ^t	2 h	79
9	6i	^t BuCH=CH-	SiMe ₂ Bu ^t	3 h	73
10	6j	PhCH=CH-	SiMe ₂ Bu ^t	4 h	46
11	6k	3-MeOC ₆ H ₄ CH=CH-	SiMe ₂ Bu ^t	21 h	62
12	61	2-ClC ₆ H ₄ CH=CH-	SiMe ₂ Bu ^t	3.5 h	71

^aNMR spectroscopies indicated the presence of only one isomer in every case, double-bond geometries of which were not determined. ^bThe reaction was performed using $T_{s}NH_{2}$ (4.0 equiv) and $Ti(OEt)_{4}$ (4.0 equiv) in THF.

yallenes **4** in combination with a Brook rearrangement.¹⁵ We envisaged this chemistry could be applicable to the enantioselective preparation of α -silylamines using α -silylimines that are readily available from the corresponding acylsilanes. Enantioselective MPV-type reduction of α -silylimines is unprecedented, although there is one example for ketimines using (BINOL)Al^{III}/2-propanol.^{16,17}

RESULTS AND DISCUSSION

On application of the MPV-type reduction for acylsilanes to imines, several issues were recognized: (1) less electrophilic reactivity of silylimines in comparison with silylketones and (2) possibility of the occurrence of an aza-Brook rearrangement¹⁸ in the reduction product before quenching. Taking into account the electron-withdrawing nature of a sulfonyl group capable of stabilizing the developing anion in the hydride transfer and convenient availability of starting materials, we chose *N*-tosylsilylimine **6** as a substrate, which can be prepared by the reaction of acylsilanes with tosylamine in the presence of triethylamine and TiCl₄ (Table 1).

At the outset, we examined the feasibility of hydride transfer to 6a from LDA^{14,19} and found that the reaction can proceed over a wide range of temperatures with no products resulting from an aza-Brook rearrangement being detected (Table 2).

Having demonstrated the possibility of the desired MPV-type reduction of an N-tosylsilylimine, we then proceeded to examine enantioselective variants using enantioenriched known lithium amides including 2 (Table 3, entries 1–3).

Table 2. Reduction of N-Tosylimine Using LDA

	Ph SiMe ₃ LDA	HN ^{-Ts} H Ph ^{-SiMe} ₃ 7a	
entry	<i>T</i> (°C)	у	ield (%)
1	0		54 66
2	-40		66
3	-80		73
4	-100		17 ^a

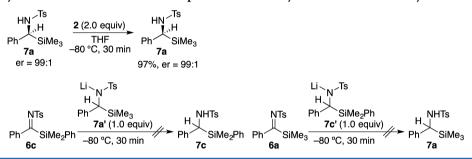
^a54% of starting material was recovered.

The best result was obtained with lithium amide 2 as in the case of acylsilanes, suggesting an important role of the nitrogen atom of piperazine. The absolute configurations of 7a-d were determined on the basis of X-ray analysis of 7a. The fact that benzaldehyde was obtained with 9 suggests that not the methyne hydrogen on the stereogenic carbon atom but the methylene hydrogen was used for hydride transfer. The inverse addition of the imine to a solution of 2 resulted in lowering the chemical yield and enantioselectivity (entry 4). The use of THF as a solvent is crucial to the success of the reaction; toluene and Et₂O resulted in recovered starting material and racemization, respectively (entries 5 and 6). The reaction proceeded with high selectivity even at 0 °C, and lowering the reaction temperature to -100 °C led to improvement in enantioselectivity to 99:1 but at the expense of chemical yield (entries 7 and 8). A prolonged reaction time or the use of additional amounts of lithium amide 2 (2.4 equiv) at -100 °C did not improve the chemical yield (entries 9 and 10). Occurrence of significant racemization with an excess amount of 2 (entries 4 and 10) indicates the possibility of deprotonation of 7a by the base. Also, the possibility of intra- and/or intermolecular proton transfer from an α -silyl carbon atom to a nitrogen atom before quenching, leading to an α -silyl benzyl carbanion, cannot be ruled out. However, the fact that enantioenriched 7a (er = 99:1) did not racemize upon treatment with 2 at -80 °C makes these possibilities unlikely (Scheme 2). While TBS derivative 6b gave the product with same enantiomeric ratio as that in the case of 6a, introduction of a phenyl group on the silyl group caused a decline in enantioselectivity. This may reflect a change in E/Z geometry of the imine moiety (vide infra) (Table 3, entries 11-13). Another possibility for the partial racemization is that the resulting lithium amide 7a' acts as a nonselective reducing agent. To test the feasibility of the process, dimethylphenylsilyl derivative 6c was treated with 7a', which was generated from 7a by deprotonation. In this reaction and the reaction using the opposite combination of trimethylsilyl and dimethylphenylsilyl groups, hydride transfer was not observed. Consequently, a decrease in enantioselectivity with an excess amount of a base might be due to other factors such as a nonselective hydride transfer from aggregated species.

		Ph		Ph SiR ₃ Ph	Me Me H I H Ph Ph B Li 9	Me N ∕ Ph Li		
entry	2, 8-10	6	SiR ₃	T (°C)	solvent	yield (%)	er	sm (%)
1	8	6a	SiMe ₃	-80	THF	0		
2	9	6a	SiMe ₃	-80	THF	65 ^b	50:50	
3	2	6a	SiMe ₃	-80	THF	76	93:7	7
4 ^{<i>c</i>}	2	6a	SiMe ₃	-80	THF	50	82:18	36
5	2	6a	SiMe ₃	-80	toluene	0		78
6	2	6a	SiMe ₃	-80	Et ₂ O	7	48:52	69
7	2	6a	SiMe ₃	0	THF	65	97:3	
8	2	6a	SiMe ₃	-100	THF	58	99:1	28
9	2	6a	SiMe ₃	-100	THF	53	93:7	19
10	2	6a	SiMe ₃	-100	THF	59	78:22	29
11	2	6b	SiMe ₂ Bu ^t	-80	THF	68	92:8	
12	2	6c	SiMe ₂ Ph	-80	THF	71	75:25	
13	2	6d	$SiMePh_2$	-80	THF	53	84:16	15

^{*a*}All reactions were performed using 1.2 equiv of lithium amide except for entry 10 (2.4 equiv) and for 30 min except for entry 9 (180 min). ^{*b*}Benzaldehyde (21%) was obtained as a byproduct. ^{*c*}Inverse addition of **6a** to a cooled solution of **2**.

Scheme 2. Possibility for Partial Racemization via Deprotonation of 7 by 2 or Reduction of 6 by Reduction Product 7'



The same type of hydride transfer reaction can be applied to α,β -unsaturated imines **6e**-**1** to give the corresponding amines **7e**-**1** in good to excellent enantioselectivity (Table 4). The absolute configurations of **7e**-**1** were determined on the basis of X-ray analysis of **7e**. In the case of β -aryl-substituted derivatives, enamine **10**, a product that can arise from the above-mentioned proton transfer followed by an allylic and then a C-to-N lithio migration, was formed as a byproduct.

Table 4. Reduction of *N*-Tosylimines Using Chiral Lihium Amide 2

NTs R 6e-	SiMe ₂ tB	u THF R	$\begin{array}{ccc} HN & TS & HN & TS \\ \overrightarrow{I} & H & \overrightarrow{I} & H \\ R & SiMe_2 Bu & Ar & Ju & SiMe_2 \\ \textbf{7e-I} & \textbf{10j-I} \end{array}$			∕le₂ ^t Bu	
					7	10	6
entry	6	R		yield (%)	er	yield (%)	yield (%)
1	6e	TMSC≡C-		91	88:12		
2	6f	PhC≡C-		84	91:9		
3	6g	$Ph(CH_2)_3C \equiv C$ -		58	97:3		
4	6h	TMSCH=CH-		90	99:1		
5	6i	^t BuCH=CH-		77	99:1		
6	6j	PhCH=CH-		67	99:1	22	
7	6k	3-MeOC ₆ H ₄ CH=CH	-	60	97:3	18	
8	61	2-ClC ₆ H ₄ CH=CH-		27	97:3	31	20

Regarding the enantioselectivity observed in the MPV-type reduction of a carbonyl group by chiral lithium amides, we have previously rationalized this by invoking a model of a chairlike six-membered transition state (early transition state), in which the silyl and the chelated piperazinylmethyl groups occupy an axial position.¹⁴ The results obtained with 6a-d and 6e-l, however, were inconsistent with those expected from the model as shown in Scheme 3. Thus, the axial/equatorial preference of the silvl group is reversed between chairlike transition states 11a and 11e, leading to (S)-7a and (R)-7e, respectively. The results indicate that in the chairlike model, phenyl and tertbutyldimethylsilyl groups must each occupy an axial position, respectively. Considering results showing that the extents of the enantioselectivities observed with 7a,b and 7e-l are almost the same despite differences in the relative sizes of the two substituents in the imine carbon atom (Ph vs SiMe₃ and RC \equiv C vs TBS), the chairlike model does not seem appropriate. Also, a distinct structural difference between N-tosyl imines and acylsilanes or ketones that potentially affects the enantioselectivity should be the presence of E/Z geometry concerning the N-tosyl group in imines. The geometries of E- and Z-6a and **6e** were optimized at the B3LYP level with the 6-31G(d) basis set, and it was found that E-isomers were more stable than the Z-isomer (6a, 0.54 kcal/mol; 6e, 3.81 kcal/mol). There have been some reports^{20,21} suggesting that N-sulfonylimines are configurationally unstable even at low temperatures, and in fact, there was no peak separation in each case when ¹H NMR of **6a** and that of 6e were measured at room temperature. The fact Scheme 3. Chair-Like Six-Membered Transition State Models of the Reduction

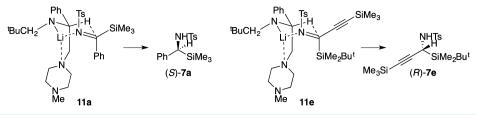
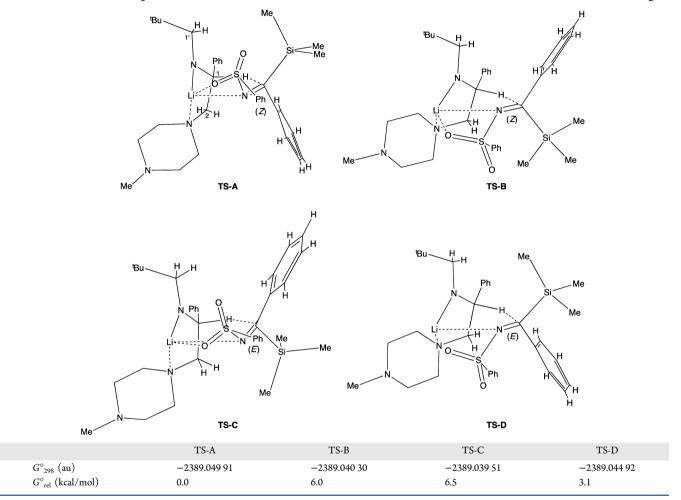


Table 5. B3LYP/6-31G* Optimized Geometries of Transition-State Structures of Reduction of 12 and Their relative Energies

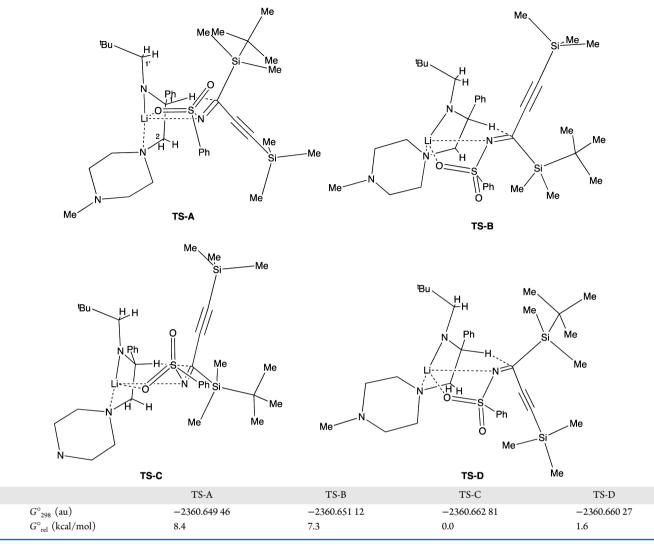


that the degree of enantioselectivity is not affected by the reaction temperature (Table 3, entries 3, 7, and 8) also indicates that the configuration of N-tosylimine in the ground state is not an important factor to control the enantioselectivity.

To obtain information on the origin of the observed enantioselectivities, we performed density functional theory (DFT) calculations on transition states leading to (S)-7a and (R)-7a using a N-phenylsulfonyl derivative 12 as a model system, assuming that the reduction proceeds by a concerted hydride transfer in a six-membered ring transition state.²² Two transition structures for each, corresponding to the (Z)- and (E)-imines in a ground state, TS-A-TS-D, were located at a B3LYP/6-31+G* level. These calculations account for the enantioselectivity observed by showing that TS-A leading to (S)-7a is the most stable (Table S).²³ The fact that the conformation of the imine in the most stable TS-A is Z suggests that E-to-Z isomerization can occur under the reaction conditions. Apparent differences between the calculated structures and the previously proposed ones in Scheme 3 are deformation from a chairlike six-membered ring in which the lithium atom coordinates to the sulfone oxygen atom rather than the imine nitrogen and increase in the double-bond character of the C1-nitrogen bond. The latter is likely to be a major factor in determining the enantioselectivity because it causes a steric interaction between the 1'-methylene group and the imine-carbon substituents. Thus, the selectivity results from a balance of steric repulsions of the imine carbon substituents with C2 and C1'.

A similar analysis can be applied to 6e (*N*-phenylsulfonyl derivative 13), in which the two substituents on the imine carbon atom are different in their bulkiness in comparison with those for 6a (Table 6). Thus, in TS-C leading to 7e, calculated to be the most stable, a much bulkier TBS group is preferably disposed at a position to avoid steric repulsions with 1'-methylene and the Ts group occupies the same side as the much less bulky silylethynyl group.

Table 6. B3LYP/6-31G* Optimized Geometries of Transition-State Structures of Reduction of 13 and Their Relative Energies



We have developed a new and efficient method for the enantioselective formation of α -silylamine by Meerwein–Ponndorf–Verley-type reduction of *N*-tosylsilylimines with lithium amide **2**. A rationale for the excellent enantioselectivity of the hydride transfer was offered on the basis of analysis of transition state models obtained from DFT calculations.

EXPERIMENTAL SECTION

General Methods. All moisture-sensitive reactions were performed under a positive pressure of nitrogen. Anhydrous MgSO₄ was used for drying all organic solvent extracts in work-up unless otherwise indicated, and removal of the solvents was performed with a rotary evaporator. Dry solvents and reagents were obtained by using standard procedures. Thin-layer chromatography was performed on precoated glass-backed silica gel 60 F-254 plates. For routine chromatography, the following adsorbents were used: silica gel 60N of particle size 63-210 μ m or 40–50 μ m. Liquid chromatography under medium pressures (MPLC) was carried out using prepacked columns (22 mm × 100 mm (5 μ m silica gel) or 22 mm × 300 mm (10 μ m silica gel)). ¹H NMR spectra (500 MHz) were taken in CDCl₃ with reference to CHCl₃ (δ 7.26). ¹³C NMR spectra (125 MHz) were measured in CDCl₃ with reference to CHCl₃ (δ 77.2). The assignment of ¹H and ¹³C NMR spectra was based on H–H decoupling and HMQC experiments. Mass spectra (HRMS) were obtained on an Orbitrap mass spectrometer.

Preparation of Acylsilanes. Compounds $5a_{,}^{24} 5b_{,}^{25} 5c_{,}^{26} 5e_{,}^{27} 5f_{,}^{28} 5g_{,}^{15} 5h_{,}^{29} 5i_{,}^{30} 5j_{,}^{31}$ and $5k^{32}$ were prepared according to literature procedures.

Diphenylmethylbenzoylsilane (5d).³¹ To a cooled (-80 °C) solution of 2-phenyl-1,3-dithiane (934 mg, 4.76 mmol) in THF (20 mL) was added a solution of n-BuLi (1.83 M in hexane, 2.86 mL, 5.23 mmol). After 2 h of stirring at the same temperature, a solution of Ph₂MeSiCl (1.2 mL, 1.33 g, 5.71 mmol) in THF (6 mL) was added. After being allowed to warm to 0 °C over 30 min, the reaction mixture was diluted with saturated aqueous NH4Cl solution (30 mL) and extracted with Et_2O (30 mL \times 3). The combined organic phases were washed with saturated brine (30 mL), dried, and concentrated. The residual oil was subjected to column chromatography (silica gel, 50 g; elution with hexane/ $CH_2Cl_2 = 4:1$) to give methyldiphenyl(2-phenyl-1,3-dithian-2-yl)silane (1.77 g, 95%). To a cooled (ice-water) solution of the above compound (807 mg, 2.06 mmol) in acetone (21 mL) and H_2O (630 μ L) was added NBS (1.83 g, 10.3 mmol). After being stirred at room temperature for 30 min, the reaction mixture was diluted with 15% aqueous Na₂S₂O₃ (10 mL) and extracted with Et₂O (30 mL \times 3). The combined organic phases were washed with H₂O $(30 \text{ mL} \times 3)$ and saturated brine (30 mL), dried, and concentrated. The residual oil was subjected to column chromatography (silica gel, 10 g; elution with hexane/Et₂O = 20:1) to give 5d (164 mg, 26%).

1-(*tert*-Butyldimethylsilyl)-3-(trimethylsilyl)prop-2-yn-1-one (5e). To a cooled (ice–water) solution of 1-(*tert*-butyldimethylsilyl)-3-

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(trimethylsilyl)prop-2-yn-1-ol³² (568 mg, 2.34 mmol) in acetone (9.4 mL) was added Jones reagent (1.94 M, 1.45 mL, 2.81 mmol). After 10 min of stirring at the same temperature, i-PrOH (1 mL) was added to the reaction mixture, which was diluted with saturated aqueous NaHCO₃ solution (10 mL) and extracted with Et₂O (10 mL \times 3). The combined organic phases were washed with H_2O (10 mL \times 3) and saturated brine (10 mL), dried, and concentrated. The residual oil was subjected to column chromatography (silica gel, 20 g, elution with hexane/Et₂O = 30:1) to give **5e** (479 mg, 85%) as an orange oil. R_f = 0.52 (hexane/Et₂O = 3:1); IR (NaCl) 2953, 2931, 2889, 2858, 1637, 1581 cm⁻¹; ¹H NMR (CDCl₂) δ 0.34 (6H, s), 0.98 (9H, s), 6.84 (1H, d, J = 16.5 Hz), 7.28–7.34 (2H, m), 7.43 (1H, dd, J = 7.8, 1.2 Hz), 7.65 (1H, dd, J = 7.6 Hz, 2.1), 7.88 (1H, d, J = 16.5 Hz); ¹³C NMR $(CDCl_2) \delta = 5.5, 16.9, 26.8, 127.3, 127.3, 130.3, 131.3, 133.3, 134.6,$ 135.6, 139.0, 236.1; HRMS-APCI (m/z) $[M + H]^+$ calcd for C15H21OClSi 281.1123, found 281.1126.

(E)-1-(tert-Butyldimethylsilyl)-3-(2-chlorophenyl)prop-2-en-1-one (51). To a cooled (ice-water) suspension of NaH (60%, 157 mg, 3.91 mmol) in THF (14 mL) was added a solution of HWE reagent (1.14 g, 4.27 mmol) in THF (13 mL). The reaction mixture was allowed to warm to room temperature and was stirred for 10 min. The mixture was cooled to 0 °C, and then 2-chlorobenzaldehyde (0.40 mL, 3.56 mmol) was added. After being stirred at room temparature for 1.5 h, the reaction mixture was diluted with saturated aqueous NH_4Cl solution (30 mL) and extracted with Et₂O (30 mL × 3). The combined organic phases were washed with saturated brine (30 mL), dried, and concentrated. The residual oil was subjected to column chromatography (silica gel, 50 g; elution with hexane/AcOEt = 15:1) to give 51 (957 mg, 96%) as an orange oil. $R_f = 0.52$ (hexane/Et₂O = 3:1); IR (NaCl) 2953, 2931, 2889, 2858, 1637, 1581 cm⁻¹; ¹H NMR $(CDCl_3) \delta 0.34 (6H, s), 0.98 (9H, s), 6.84 (1H, d, J = 16.5 Hz), 7.28-$ 7.34 (2H, m), 7.43 (1H, dd, J = 7.8, 1.2 Hz), 7.65 (1H, dd, J = 7.6 Hz, 2.1), 7.88 (1H, d, J = 16.5 Hz); ¹³C NMR (CDCl₃) δ -5.5, 16.9, 26.8, 127.3, 127.3, 130.3, 131.3, 133.3, 134.6, 135.6, 139.0, 236.1; HRMS-APCI (m/z) [M + H]⁺ calcd for C₁₅H₂₁OClSi 281.1123, found 281.1126.

Representative Procedure for the Preparation of *N*-Silylimine (6i). To a cooled solution of acylsilane (325 mg, 1.44 mmol), TsNH₂ (294 mg, 1.72 mmol), and Et₃N (881 μ L, 6.32 mmol) in CH₂Cl₂ (7.2 mL) was added TiCl₄ (1.0 M in CH₂Cl₂, 1.44 mL, 1.44 mmol). After 3 h of stirring at room temperature, saturated aqueous NaHCO₃ (10 mL) was added. The mixture was filtered through a plug of Celite, and the filtrate was extracted with CH₂Cl₂ (20 mL × 2). The combined organic phases were washed with H₂O (10 mL) and saturated brine (10 mL), dried, and concentrated. The residual oil was subjected to column chromatography (silica gel, 30 g; elution with hexane/CH₂Cl₂/Et₂O = 40:20:1) to give 6i (397 mg, 73%).

Preparation of N-Silylimine (6g). To a solution of asylsilane (493 mg, 1.72 mmol) and TsNH₂ (1.18 g, 6.88 mmol) in THF (5.7 mL) was added Ti(OEt)₄ (1.44 mL, 1.57 g, 6.88 mmol). The mixture was heated to reflux for 8 h. After cooling to room temperature, saturated aqueous NaHCO₃ (10 mL) was added. The mixture was filtered through a plug of Celite, and the filtrate was extracted with AcOEt (10 mL \times 2). The combined organic phases were dried and concentrated. After the residue was passed through a pad of silica gel eluting with hexane/CH₂Cl₂/Et₂O = 5:5:1, the residual oil was subjected to column chromatography (silica gel, 30 g; elution with hexane/Et₂O = 5:1) to give **6g** (419 mg, 55%)

N-(Phenyl(trimethylsilyl)methylene)-*p*-toluenesulfonamide (6a). Colorless needles (CH₂Cl₂/hexane). 67% yield, 2.45 g (from 1.99 g of 5a); mp = 112.0–113.0 °C; R_f = 0.43 (hexane/AcOEt = 5:1); IR (KBr) 3358, 3260, 2965, 1740, 1550 cm⁻¹; ¹H NMR (CDCl₃) δ 0.07–0.41 (9H, brs), 2.40 (3H, s), 7.14–7.29 (4H, brm), 7.33–7.29 (3H, m), 7.71–7.82 (2H, br); ¹³C NMR (CDCl₃) δ –2.2 (br), 21.7, 125.0 (br), 127.7, 128.2, 129.2–129.8 (br), 205.8; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₁₇H₂₁NO₂SSi 354.0955, found 354.0957.

N-(Phenyl(trimethylsilyl)methylene)-*p*-toluenesulfonamide (**6b**). Colorless needles (CH₂Cl₂/hexane). 74% yield, 634 mg (from 508 mg of **5b**); mp = 69.5–70.0 °C; R_f = 0.48 (hexane/AcOEt = 5:1); IR (KBr) 3055, 2959, 2927, 2890, 2856, 1597, 1548; ¹H NMR (CDCl₃) δ 0.14 (6H, s), 0.90 (9H, s), 2.41 (3H, s), 7.18–7.20 (2H, m), 7.24 (2H, d, *J* = 8.3 Hz), 7.35–7.39 (3H, m), 7.74 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –5.8, 17.9, 21.7, 26.8, 125.1, 127.6, 128.2, 129.2, 129.4, 138.6, 140.8, 143.5, 205.3; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₀H₂₇NO₂SSi 396.1424, found 396.1427.

N-((Methyldiphenylsilyl)(phenyl)methylene)-*p*-toluenesulfonamide (6d). Pale yellow oil. 56% yield, 133 mg (from 164 mg of 5d); $R_f = 0.44$ (hexane/Et₂O = 3:2); IR (NaCl) 3065, 3024, 2965, 2922, 1595, 1551 cm⁻¹; ¹H NMR (CDCl₃) δ 0.74 (1H, brs), 2.42 (3H, s), 7.07 (2H, d, J = 7.4 Hz), 7.21–7.26 (4H, m), 7.28–7.35 (5H, m), 7.40–7.44 (2H, m), 7.48 (4H, d, J = 7.1 Hz), 7.72 (2H, d, J = 8.2 Hz); ¹³C NMR (CDCl₃) δ -4.1, 21.7, 125.9, 127.7, 128.0, 128.2, 129.5, 129.6, 130.3, 132.5, 135.4, 138.3, 143.6, 202.0; HRMS-ESI (m/z) [M + Na]⁺ calcd for C₂₇H₂₅NO₂SSi 478.1268, found 478.1267.

N-(1-(*tert*-Butyldimethylsilyl)-3-(trimethylsilyl)prop-2-yn-1ylidene)-*p*-toluenesulfonamide (6e). Off-white plates (CH₂Cl₂/ hexane). 97% yield, 852 mg (from 538 mg of 5e); mp = 65.0–66.0 °C; $R_f = 0.34$ (hexane/Et₂O = 3:1); IR (KBr) 2967, 2930, 2897, 2859, 1596, 1555; ¹H NMR (CDCl₃) d 0.22 (s, 6H), 0.24 (9H, s), 0.91 (9H, s), 2.43 (3H, s), 7.29 (2H, d, *J* = 8.3 Hz), 7.85 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –6.7, –0.7, 17.9, 21.8, 26.6, 100.5, 128.1, 128.3, 129.5, 137.1, 144.0, 184.6; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₁₉H₃₁NO₂SSi₂ 416.1506, found 416.1500.

N-(1-(*tert*-Butyldimethylsilyl)-3-phenylprop-2-yn-1-ylidene)*p*-toluenesulfonamide (6f). Pale yellow needles (CH₂Cl₂/hexane). 83% yield, 366 mg (from 270 mg of 5f); mp = 70.5–71.0 °C; R_f = 0.50 (hexane/Et₂O = 2:1); IR (KBr) 2948, 2929, 2897, 2855, 2170, 1509 cm⁻¹; ¹H NMR (CDCl₃) δ 0.30 (6H, s), 0.96 (9H, s), 2.41 (3H, s), 7.30 (2H, d, *J* = 8.3 Hz), 7.41 (2H, dd, *J* = 7.7, 7.7 Hz), 7.47 (1H, tt, 7.7, 1.2 Hz), 7.57 (2H, dd, *J* = 7.7, 1.2 Hz), 7.90 (1H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –6.6, 18.0, 21.8, 26.6, 87.7, 118.3, 121.2, 128.1, 128.8, 129.6, 131.3, 133.0, 137.3, 144.0, 183.5; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₂₇NO₂SSi 420.1424, found 420.1421.

N-(1-(*tert*-Butyldimethylsilyl)-6-phenylhex-2-yn-1-ylidene)*p*-toluenesulfonamide (6g). Pale yellow oil. $R_f = 0.29$ (hexane/ Et₂O = 5:1); IR (NaCl) 3061, 3027, 2952, 2931, 2893, 2859, 2189, 1600, 1515 cm⁻¹; ¹H NMR (CDCl₃) δ 0.24 (6H, s), 0.94 (9H, s), 1.93 (2H, tt, *J* = 7.1. 7.6 Hz), 2.41 (3H, s), 2.57 (2H, t, *J* = 7.1 Hz), 2.77 (2H, t, *J* = 7.6 Hz), 7.19–7.22 (3H, m), 7.28–7.32 (4H, m), 7.86 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –6.7, 17.7, 20.2, 21.7, 26.5, 29.6, 34.9, 80.6, 122.1, 126.2, 127.9, 128.6, 128.7, 129.4, 137.6, 141.1, 143.7; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₅H₃₃NO₂SSi 462.1894 found 462.1894.

N-(2*E*)-1-(*tert*-Butyldimethylsilyl)-3-(trimethylsilyl)allylidene)-*p*-toluenesulfonamide (6h). Orange oil. 79% yield, 877 mg (from 682 mg of 5h); $R_f = 0.47$ (hexane/Et₂O = 5:1); IR (NaCl) 2956, 2931, 2894, 2859, 1599, 1515; ¹H NMR (CDCl₃) δ 0.17 (9H, s), 0.21 (6H, s), 0.89 (9H, s), 2.43 (3H, s), 6.64 (1H, d, *J* = 19.5 Hz), 7.56 (1H, d, *J* = 19.5 Hz), 7.30 (2H, d, *J* = 8.3 Hz), 7.84 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ -4.3, -1.7, 17.6, 21.7, 26.9, 127.3, 129.5, 139.2, 142.2, 143.4, 149.8, 199.8; HRMS-ESI (*m*/*z*); [M + Na]⁺ calcd for C₁₉H₃₃NO₂SSi₂ 418.1663, found 418.1659.

N-((*E*)-1-(*tert*-Butyldimethylsilyl)-4,4-dimethylpent-2-en-1ylidene)-*p*-toluenesulfonamide (6i). Pale yellow oil. $R_f = 0.28$ (hexane/Et₂O = 8:1); IR (NaCl) 2959, 2932, 2860, 1623, 1513 cm⁻¹; ¹H NMR (CDCl₃) d 0.23 (6H, s), 0.89 (9H, s), 1.12 (9H, s), 6.58 (1H, d, *J* = 16.1 Hz), 7.24 (1H, d, *J* = 16.1 Hz), 7.29 (2H, d, *J* = 8.3 Hz), 7.84 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) d -3.8, 17.5, 21.7, 27.0, 28.7, 35.0, 126.3, 127.2, 129.5, 139.5, 143.2, 162.2, 198.7; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₀H₃₃NO₂SSi 402.1894, found 402.1898.

N-((*E*)-1-(*tert*-Butyldimethylsilyl)-3-phenylallylidene)-*p*-toluenesulfonamide (6j). Pale yellow needles (CH₂Cl₂/hexane). 46% yield, 512 mg (from 687 mg of 5j); mp = 105.0–105.5 °C; R_f = 0.39 (hexane/Et₂O = 3:1); IR (KBr) 2952, 2928, 2887, 2857, 1613 cm⁻¹; ¹H NMR (CDCl₃) δ 0.32 (6H, s), 0.94 (9H, s), 2.43 (3H, s), 7.29–7.34 (3H, m), 7.41–7.42 (3H, m), 7.58–7.59 (2H, m), 8.07 (1H, d, *J* = 16.3 Hz); ¹³C NMR (CDCl₃) δ –3.7, 17.7, 21.7, 27.0, 127.1, 127.6, 128.7, 129.2, 129.5, 131.1, 135.0, 139.3, 143.3, 147.3, 197.1; HRMS-

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ESI (m/z) [M + Na]⁺ calcd for C₂₂H₂₉NO₂SSi 422.1581, found 422.1583.

N-((*E*)-1-(*tert*-Butyldimethylsilyl)-3-(3-methoxyphenyl)allylidene)-*p*-toluenesulfonamide (6k). Yellow prisms (CH₂Cl₂/ hexane). 62% yield, 462 mg (from 482 mg of 5k); mp = 90.0−91.0 °C; $R_f = 0.30$ (hexane/Et₂O = 3:1); IR (KBr) 2958, 2926, 2857, 1615, 1588 cm⁻¹; ¹H NMR (CDCl₃) δ 0.32 (6H, s), 0.94 (9H, s), 2.40 (3H, s), 3.83 (3H, s), 6.95 (1H, dd, *J* = 8.3, 1.6 Hz), 7.08 (1H, brs), 7.17 (1H, d, *J* = 7.6 Hz), 7.29−7.32 (4H, m), 7.88 (2H, d, *J* = 8.2 Hz), 8.05 (1H, d, *J* = 16.3 Hz); ¹³C NMR (CDCl₃) δ −3.9, 17.6, 21.6, 26.9, 55.4, 113.3, 116.8, 121.3, 127.0, 127.7, 129.4, 130.1, 136.3, 139.2, 143.3, 147.0, 160.1, 197.0; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₃H₃₁NO₃SSi 452.1686, found 452.1687.

N-((*E*)-1-(*tert*-Butyldimethylsilyl)-3-(2-chlorophenyl)allylidene)-*p*-toluenesulfonamide (6l). Yellow prisms (CH₂Cl₂/ hexane). 71% yield, 662 mg (from 606 mg of **5**l); mp = 113.0–114.0 °C; R_f = 0.33 (hexane/Et₂O = 3:1); IR (KBr) 2952, 2926, 2885, 2853, 1611, 1503 cm⁻¹; ¹H NMR (CDCl₃) δ 0.35 (6H, s), 0.95 (9H, s), 2.41 (3H, s), 7.29–7.33 (4H, m), 7.39–7.43 (1H, m), 7.78 (1H, d, *J* = 16.3 Hz), 7.78–7.80 (1H, m), 7.89 (2H, d, *J* = 8.0 Hz), 8.07 (1H, d, *J* = 16.3 Hz); ¹³C NMR (CDCl₃) δ −4.0, 17.6, 21.7, 26.9, 127.1, 127.5, 127.9, 129.1, 129.5, 130.1, 131.7, 133.1, 135.4, 139.1, 142.4, 143.4, 197.2; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₂₈NO₂ClSSi 456.1191, found 456.1190.

Representative Procedure for the Reduction of *N***-Silylimine (6b).** To a cooled $(-80 \ ^{\circ}C)$ solution of **6b** (92.7 mg, 0.248 mmol) in THF (1.2 mL) was added dropwise a solution of chiral lithium amide generated from (*S*)-2,2-dimethyl-*N*-(2-(4-methylpiperazin-1-yl)-1-phenylethyl)propan-1-amine (83.4 mg, 0.288 mmol) and *n*-BuLi (1.97 M in *n*-hexane, 151 μ L, 0.298 mmol) in THF (1.1 mL) at 0 $^{\circ}C$. The reaction mixture was stirred at the same temperature for 30 min before the addition of a solution of AcOH (0.5 M in THF, 0.600 mmol). The mixture was diluted with hydrochloric acid (1%, 10 mL) and extracted with AcOEt (10 mL × 3). The combined organic phases were successively washed with saturated aqueous NaHCO₃ solution (5 mL) and saturated brine (5 mL), dried, and concentrated. The residual oil was subjected to flash column chromatography (silica gel, 7 g, elution with hexane/CH₂Cl₂/Et₂O = 20:20:1) to give 7b (54.1 mg, 68%).

N-(Phenyl(trimethylsilyl)methyl)-*p*-toluenesulfonamide (7a). Colorless needles (CH₂Cl₂/hexane). 58% yield, 38 mg (from 73 mg of **6a**); mp = 170–171 °C; $R_f = 0.34$ (hexane/AcOEt = 5:1); $[\alpha]^{27}_{\rm D}$ -60.0 (*c* 0.35, CHCl₃); Chiralcel OD-H (25 cm), hexane/*i*-PrOH = 10:1, flow rate 1.0 mL/min, detection at 254 nm, $t_{\rm R} = 7.91$ min (major) and 11.6 min (minor), er = >99:1; IR (KBr) 3284, 2964, 1598; ¹H NMR (CDCl₃) δ -0.02 (9H, s), 2.29 (3H, s), 3.97 (1H, d, *J* = 8.4 Hz), 5.23 (1H, d, *J* = 8.4 Hz), 6.81 (2H, d, *J* = 7.1 Hz), 6.97–7.05 (5H, m), 7.49 (2H, d, *J* = 8.2 Hz); ¹³C NMR (CDCl₃) δ -3.7, 21.5, 50.4, 125.6, 126.3, 127.4, 128.1, 129.2, 137.7, 139.4, 143.0; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₁₇H₂₃O₂NSSi 356.1111, found 356.1114.

N-((*tert*-Butyldimethylsilyl)(phenyl)methyl)-*p*-toluenesulfonamide (7b). Colorless prisms (CH₂Cl₂/hexane); mp = 135.5–136.5 °C; $R_f = 0.35$ (hexane/AcOEt = 5:1); $[\alpha]^{26}_{D} - 23.0$ (*c* 1.02, CHCl₃) (er = 93:7); Chiralcel OD-H (25 cm), hexane/*i*-PrOH = 8:1, flow rate 1.00 mL/min, detection at 254 nm, $t_R = 5.99$ min (major) and 7.22 min (minor), er = 93:7; IR (KBr) 3476, 3304, 2954, 2927, 2886, 2855, 1599 cm⁻¹; ¹H NMR (CDCl₃) δ -0.29 (3H, s), 0.05 (3H, s), 0.90 (9H, s), 2.26 (3H, s), 4.19 (1H, d, *J* = 8.9 Hz), 4.98–5.05 (1H, brm), 6.78 (2H, d, *J* = 6.9 Hz), 6.92–7.00 (5H, m), 7.41 (2H, d, *J* = 8.0 Hz); ¹³C NMR (CDCl₃) δ -8.5, -7.0, 17.5, 21.5, 26.9, 48.0, 125.6, 126.7, 127.4, 128.1, 129.1, 137.9, 139.9, 142.8; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₀H₂₉NO₂SSi 398.1581, found 398.1585.

N-((Dimethyl(phenyl)silyl)(phenyl)methyl)-*p*-toluenesulfonamide (7c). Colorless needles (AcOEt/hexane). 71% yield, 41 mg (from 66 mg of 6c); mp = 151.0–152.0 °C; R_f = 0.33 (hexane/AcOEt = 4:1); [α]²⁵_D –25.9 (c 1.03, CHCl₃) (er = 68:32); Chiralcel OD-H (25 cm), hexane/*i*-PrOH = 10:1, flow rate 1.00 mL/min, detection at 254 nm, t_R = 7.96 min (major) and 21.9 min (minor), er = 75:25; IR (KBr) 3267, 3063, 3026, 2957, 1595 cm⁻¹; ¹H NMR (CDCl₃) δ 0.24 (3H, s), 0.27 (3H, s), 2.29 (3H, s), 4.14 (1H, d, *J* = 8.3 Hz), 5.26–5.29 (1H, m), 6.72–6.73 (2H, m), 6.96 (2H, d, *J* = 8.1 Hz), 6.99–7.02 (3H, m), 7.28–7.31 (4H, m), 7.37–7.42 (1H, m), 7.41 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –5.6, –4.8, 21.5, 49.9, 125.6, 126.5, 127.3, 127.9, 128.1, 129.1, 130.0, 134.1, 134.5, 137.4, 138.9, 142.8 ; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₂₅NO₂SSi 418.1268, found 418.1261.

N-((Methyldiphenylsilyl)(phenyl)methyl)-*p*-toluenesulfonamide (7d). White prisms (CH₂Cl₂/hexane). 53% yield, 39 mg (from 73 mg of 6d); mp = 174.0–175.0 °C; R_f = 0.34 (hexane/Et₂O = 3:2); [α]²⁷_D -23.8 (*c* 0.72, CHCl₃) (er = 84:16); Chiralpak AD-H (25 cm), hexane/*i*-PrOH = 15:1, flow rate 1.00 mL/min, detection at 254 nm, t_R = 10.3 min (major) and 15.8 min (minor), er = 84:16; IR (KBr) 3266, 3063, 3022, 2957, 2918, 1597 cm⁻¹; ¹H NMR (CDCl₃) δ 0.39 (3H, s), 2.29 (3H, s), 4.57 (1H, d, *J* = 8.0 Hz), 4.79 (1H, d, *J* = 8.0 Hz), 6.65 (2H, d, *J* = 6.7 Hz), 6.94–7.00 (5H, m), 7.30–7.47 (12H, m); ¹³C NMR (CDCl₃) δ –5.6, 21.5, 48.5, 125.9, 126.9, 127.5, 127.9, 128.2, 128.4, 129.2, 130.2, 130.5, 132.1, 133.0, 135.2, 135.6, 137.5, 138.5, 143.0; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₇H₂₇NO₂SSi 480.1424, found 480.1424.

N-(1-(*tert*-Butyldimethylsilyl)-3-(trimethylsilyl)prop-2-yn-1yl)-*p*-toluenesulfonamide (7e). Colorless plates (CH₂Cl₂/hexane). 91% yield, 72 mg (from 85 mg of 6e); mp = 161.0–161.5 °C; R_f = 0.50 (hexane/Et₂O = 2:1); $[\alpha]^{26}_{D}$ +70.9 (*c* 1.03, CHCl₃) (er = 95:5); Chiralpak AS-3 (25 cm), hexane/*i*-PrOH/EtOH = 30:1:1, flow rate 0.40 mL/min, detection at 254 nm, t_R = 11.4 min (major) and 13.6 min (minor), er = 95:5; IR (KBr) 3279, 2955, 2935, 2861, 2161, 1734, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ –0.08 (9H, s), 0.09 (3H, s), 0.11 (3H, s), 0.99 (9H, s), 2.42 (3H, s), 3.80 (1H, d, *J* = 10.5 Hz), 4.15 (1H, d, *J* = 10.5 Hz), 7.28 (2H, d, *J* = 8.1 Hz), 7.80 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –8.0, –7.5, –0.2, 17.6, 21.7, 27.0, 35.3, 90.3, 103.6, 128.1, 129.7, 137.6, 143.5; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₁₉H₃₃NO₂SSi₂ 418.1663, found 418.1656.

N-(1-(*tert*-Butyldimethylsilyl)-3-phenylprop-2-yn-1-yl)-*p*-toluenesulfonamide (7f). Colorless needles (CH₂Cl₂/hexane). 84% yield, 65 mg (from 84 mg of 6f); mp = 135.0–136.0 °C; R_f = 0.44 (hexane/Et₂O = 2:1); [α]²⁶_D +118.8 (*c* 1.08, CHCl₃) (er = 90:10); Chiralcel OD-H (25 cm), hexane/*i*-PrOH = 12:1, flow rate 1.00 mL/min, detection at 254 nm, t_R = 5.46 min (minor) and 7.29 min (major), er = 90:10; IR (KBr) 3290, 3060, 2926, 2853, 1599 cm⁻¹; ¹H NMR (CDCl₃) δ 0.15 (3H, s), 0.17 (3H, s), 1.03 (9H, s), 2.31 (3H, s), 4.01 (1H, d, *J* = 10.7 Hz), 4.27 (1H, d, *J* = 10.7 Hz), 6.93 (2H, d, *J* = 8.3 Hz), 7.17–7.25 (5H, m), 7.84 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –7.9, –7.3, 17.6, 21.6, 27.0, 34.8, 85.6, 87.2, 123.2, 128.1, 129.7, 129.7, 131.4, 131.4, 137.7, 143.7; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₂₉NO₂SSi 422.1581, found 422.1574.

N-(1-(*tert*-Butyldimethylsilyl)-6-phenylhex-2-yn-1-yl)-*p*-toluenesulfonamide (7g). Colorless prisms (CH₂Cl₂/hexane). 58% yield, 43 mg (from 78 mg of 6g); mp = 82.5–83.5 °C; R_f = 0.50 (hexane/Et₂O = 2:1); $[\alpha]^{25}_{\rm D}$ +71.3 (*c* 1.08, CHCl₃) (er = 97:3); Chiralcel OD-H (25 cm), hexane/*i*-PrOH = 10:1, flow rate 1.0 mL/min, detection at 254 nm, $t_{\rm R}$ = 5.69 min (minor) and 9.38 min (major), er = 97:3; IR (KBr) 3254, 2951, 2931, 2900, 2854, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ 0.08 (3H, s), 0.11 (3H, s), 0.99 (9H, s), 1.49 (2H, tt, *J* = 7.2, 7.2 Hz), 1.82 (2H, td, *J* = 7.2, 2.3 Hz), 2.31 (3H, s), 2.48 (2H, t, *J* = 7.2 Hz), 3.78 (1H, dd, *J* = 10.6, 2.3 Hz), 4.20 (1H, d, *J* = 10.6 Hz), 7.09 (2H, d, *J* = 7.3 Hz), 7.18–7.29 (5H, m), 7.80 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ -8.0, -7.3, 17.4, 18.4, 21.6, 27.0, 30.3, 34.5, 35.0, 78.1, 85.6, 126.2, 128.1, 128.5, 129.4, 129.5, 137.9, 141,7, 143.3; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₅H₃₅NO₂SSi 464.2050, found 464.2048.

(*E*)-*N*-(1-(*tert*-Butyldimethylsilyl)-3-(trimethylsilyl)allyl)-*p*-toluenesulfonamide (7h). Colorless prisms (CH₂Cl₂/hexane). 90% yield, 69 mg (from 79 mg of 6h); mp = 136.0–137.0 °C; R_f = 0.22 (hexane/Et₂O = 5:1); $[\alpha]^{24}_{\rm D}$ +17.5 (*c* 1.03, CHCl₃) (er = >99:1); Chiralcel OD-H (25 + 25 cm), hexane/*i*-PrOH = 25:1, flow rate 0.30 mL/min, detection at 254 nm, $t_{\rm R}$ = 34.7 min (minor) and 36.3 min (major), er = >99:1; IR (KBr) 3296, 2958, 2930, 2859, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ –0.19 (9H, s), –0.09 (3H, s), –0.02 (3H, s), 0.93 (9H, s), 2.37 (3H, s), 3.77–3.80 (1H, m), 4.84–4.89 (1H, brm), 5.19

(1H, dd, *J* = 18.9, 1.2 Hz), 5.58 (1H, dd, *J* = 18.9, 6.7 Hz), 7.20 (2H, d, *J* = 8.3 Hz), 7.71 (2H, d, *J* = 8.3 Hz); 13 C NMR (CDCl₃) δ –8.2, –7.2, –1.3, 17.6, 21.6, 27.1, 49.2, 126.7, 127.7, 129.5, 138.6, 143.1, 143.3; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₁₉H₃₅NO₂SSi₂ 420.1819, found 420.1813.

(*E*)-*N*-(1-(*tert*-Butyldimethylsilyl)-4,4-dimethylpent-2-en-1yl)-*p*-toluenenesulfonamide (7i). Colorless prisms (CH₂Cl₂/ hexane). 77% yield, 63 mg (from 88 mg of 6i); mp = 136.0–137.0 °C; $R_f = 0.32$ (hexane/Et₂O = 3:1); $[\alpha]^{27}_{D} - 18.9$ (*c* 1.12, CHCl₃) (er = 94:6); Chiralcel OD-H (25 + 25 cm), hexane/*i*-PrOH = 20:1, flow rate 0.50 mL/min, detection at 254 nm, $t_R = 19.7$ min (minor) and 20.8 min (major), er = 99:1; IR (KBr) 3299, 2959, 2830, 2859, 1599 cm⁻¹; ¹H NMR (CDCl₃) δ -0.08 (3H, s), -0.01 (3H, s), 0.72 (9H, s), 0.93 (9H, s), 2.38 (3H, s), 3.64 (1H, dd, *J* = 8.3, 8.3 Hz), 4.49 (1H, brd, *J* = 8.3 Hz), 4.88 (1H, dd, *J* = 8.3, 15.7 Hz), 5.06 (1H, d, *J* = 15.7 Hz), 7.22 (2H, d, *J* = 8.3 Hz), 7.71 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ -8.1, -7.1, 17.5, 21.6, 27.2, 29.4, 32.7, 46.3, 122.7, 127.7, 129.6, 138.9, 140.2, 143.1; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₀H₃₅NO₂SSi 404.2050, found 404.2060.

(*E*)-*N*-(1-(*tert*-Butyldimethylsilyl)-3-phenylallyl)-*p*-toluenesulfonamide (7j). Colorless needles (CH₂Cl₂/hexane). 67% yield, 56 mg (from 80 mg of 6j); mp = 140.0–140.5 °C; R_f = 0.13 (hexane/ Et₂O = 3:1); [α]²⁸_D +91.7 (*c* 1.05, CHCl₃) (er = 97:3); Chiralcel OD-H (25 + 15 cm), hexane/*i*-PrOH = 20:1, flow rate 0.70 mL/min, detection at 254 nm, t_R = 18.4 min (minor) and 27.9 min (major), er = 97:3; IR (KBr) 3252, 2953, 2931, 2856, 2361, 2337, 1597 cm⁻¹; ¹H NMR (CDCl₃) δ –0.07 (3H, s), 0.06 (3H, s), 0.96 (9H, s), 2.20 (3H, s), 3.84 (1H, dd, *J* = 8.4, 8.4 Hz), 4.85–4.92 (1H, brm), 5.69 (1H, dd, *J* = 15.9, 8.4 Hz), 5.97 (1H, d, *J* = 15.9 Hz), 6.98 (2H, d, *J* = 7.3 Hz), 7.11 (2H, d, *J* = 8.1 Hz), 7.14 (1H, t, *J* = 7.3 Hz), 7.21 (2H, dd, *J* = 7.3, 7.3 Hz), 7.74 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –8.0, -7.1, 17.5, 21.4, 27.1, 46.9, 126.0, 127.0, 127.7, 128.0, 128.4, 128.4, 129.5, 137.1, 138.5, 143.4; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₃₁NO₂SSi 424.1737, found 424.1735.

(*E*)-*N*-(1-(*tert*-Butyldimethylsilyl)-3-(3-methoxyphenyl)allyl)*p*-toluenesulfonamide (7k). Colorless needles (CH₂Cl₂/hexane). 60% yield, 52 mg (from 89 mg of 6k); mp = 139.0–140.0 °C; $R_f =$ 0.16 (hexane/Et₂O = 2:1); $[\alpha]^{27}_{D} - 24.3$ (*c* 1.25, CHCl₃) (er = 97:3); Chiralcel OD-H (25 + 15 cm), hexane/*i*-PrOH = 10:1, flow rate 0.70 mL/min, detection at 254 nm, $t_R = 17.0$ min (minor) and 29.4 min (major), er = 97:3; IR (KBr) 3299, 3029, 2948, 2931, 2890, 2857, 1645, 1604, 1576 cm⁻¹; ¹H NMR (CDCl₃) δ -0.02 (3H, s), 0.04 (3H, s), 0.95 (9H, s), 2.23 (3H, s), 3.77 (3H, s), 3.84 (1H, dd, *J* = 8.3, 8.3 Hz), 4.65–4.70 (1H, brm), 5.67 (1H, dd, *J* = 15.8, 8.3 Hz), 5.93 (1H, d, *J* = 15.8 Hz), 6.50 (1H, brs), 6.58 (1H, d, *J* = 7.6 Hz), 6.70 (1H, dd, *J* = 8.3, 2.5 Hz), 7.10–7.14 (3H, m), 7.72 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ -8.0, -7.1, 17.5, 21.5, 27.1, 46.9, 55.3, 111.8, 112.3, 118.8, 127.8, 128.3, 128.4, 129.4, 129.6, 138.5, 138.6, 143.5, 159.8; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₃H₃₃NO₃SSi 454.1843, found 454.1842.

(*E*)-*N*-(1-(*tert*-Butyldimethylsilyl)-3-(2-chlorophenyl)allyl)-*p*toluenesulfonamide (7l). Colorless needles (CH₂Cl₂/hexane). 27% yield, 23 mg (from 87 mg of 6l); mp = 139.0–140.0 °C; R_f = 0.24 (hexane/Et₂O = 2:1); $[\alpha]^{26}_{\rm D}$ +66.9 (*c* 1.07, CHCl₃) (er = 97:3); Chiralcel OD-H (25 + 15 cm), hexane/*i*-PrOH = 20:1, flow rate 0.70 mL/min, detection at 254 nm, $t_{\rm R}$ = 20.5 min (minor) and 30.2 min (major), er = 97:3; IR (KBr) 3291, 2952, 2931, 2891, 2856, 1633, 1597 cm⁻¹; ¹H NMR (CDCl₃) δ 0.00 (3H, s), 0.07 (3H, s), 0.96 (9H, s), 2.22 (3H, s), 3.85 (1H, dd, *J* = 8.5, 8.5 Hz), 4.68 (1H, d, *J* = 8.5 Hz), 5.69 (1H, dd, *J* = 16.1, 8.5 Hz), 6.36 (1H, d, *J* = 16.1 Hz), 6.96– 6.98 (1H, m), 7.07–7.11 (2H, m), 7.15 (2H, d, *J* = 8.1 Hz), 7.26–7.28 (1H, m), 7.74 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –8.0, –7.1, 17.5, 21.4, 27.1, 47.1, 124.6, 126.5, 126.6, 127.8, 128.1, 129.6, 131.2, 132.5, 135.2, 138.1, 143.4; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₃₀NO₂ClSSi 458.1347, found 458.1347.

N-(1-(*tert*-Butyldimethylsilyl)-3-phenylprop-1-en-1-yl)-*p*-toluenesulfonamide (10j). Colorless plates (CH₂Cl₂/hexane). 22% yield, 18 mg (from 80 mg of 6j); mp = 96.5–97.0 °C; R_f = 0.26 (hexane/Et₂O = 3:1); IR (KBr) 3276, 2954, 2926, 2891, 2853, 1599 cm⁻¹; ¹H NMR (CDCl₃) δ 0.13 (6H, s), 0.87 (9H, s), 2.42 (3H, s), 3.38 (2H, d, *J* = 8.0 Hz), 5.56 (1H, s), 6.35 (1H, t, *J* = 8.0 Hz), 6.97 (2H, d, *J* = 7.4 Hz), 7.17–7.24 (5H, m), 7.67 (2H, d, *J* = 8.3 Hz); ¹³C NMR (CDCl₃) δ –4.2, 18.1, 21.7, 26.8, 36.2, 126.2, 128.0, 128.3, 128.5, 129.4, 129.6, 133.5, 137.0, 140.3, 143.7; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₃₁NO₂SSi 424.1737, found 424.1734.

N-(1-(*tert*-Butyldimethylsilyl)-3-(3-methoxyphenyl)prop-1en-1-yl)-*p*-toluenesulfonamide (10k). Colorless prisms (CH₂Cl₂/ hexane). 18% yield, 16 mg (from 89 mg of 6k); mp = 97.0–98.0 °C; R_f = 0.28 (hexane/Et₂O = 2:1); IR (KBr) 3282, 3060, 2961, 2926, 2884, 2852, 1908, 1598 cm⁻¹; ¹H NMR (CDCl₃) δ 0.13 (6H, s), 0.87 (9H, s), 2.41 (3, s), 3.35 (2H, d, *J* = 8.0 Hz), 3.78 (3H, s), 5.54 (1H, s), 6.33 (1H, t, *J* = 8.0 Hz), 6.57–6.58 (2H, m), 6.74 (1H, dd, *J* = 8.0, 2.3 Hz), 7.15 (1H, dd, *J* = 8.0, 8.0 Hz), 7.22 (2H, d, *J* = 8.1 Hz), 7.67 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –4.2, 18.1, 21.7, 26.8, 36.2, 55.3, 111.6, 114.1, 120.7, 127.9, 129.4, 129.5, 129.7, 133.5, 137.1, 141.9, 143.8, 159.9; HRMS-ESI (*m*/z) [M + Na]⁺ calcd for C₂₃H₃₃NO₃SSi 454.1843, found 454.1843.

N-(1-(*tert*-Butyldimethylsilyl)-3-(2-chlorophenyl)prop-1-en-1-yl)-*p*-toluenesulfonamide (10l). Colorless plates (Et₂O/hexane). 31% yield, 27 mg (from 87 mg of 6l); mp = 120.0–120.5 °C; *R_f* = 0.39 (hexane/Et₂O = 2:1); IR (KBr) 3444, 3279, 2953, 2928, 2852, 1597 cm⁻¹; ¹H NMR (CDCl₃) δ 0.12 (6H, s), 0.87 (9H, s), 2.43 (3H, s), 3.44 (2H, d, *J* = 8.0 Hz), 5.59 (1H, s), 6.30 (1H, t, *J* = 8.0 Hz), 6.91 (1H, d, *J* = 7.5 Hz), 7.10 (1H, dd, *J* = 7.5, 7.5 Hz), 7.14 (1H, dd, *J* = 7.5, 7.5 Hz), 7.26 (2H, d, *J* = 8.1 Hz), 7.31 (1H, d, *J* = 7.5 Hz), 7.72 (2H, d, *J* = 8.1 Hz); ¹³C NMR (CDCl₃) δ –4.3, 18.1, 21.8, 26.8, 33.5, 126.9, 127.6, 128.0, 129.3, 129.4, 129.7, 133.9, 134.7, 137.1, 138.1, 143.9; HRMS-ESI (*m*/*z*) [M + Na]⁺ calcd for C₂₂H₃₀O₂NClSSi 458.1347, found 458.1346.

ASSOCIATED CONTENT

S Supporting Information

Copies of ¹H and ¹³C NMR spectra of all new compounds, chiral HPLC analyses of compound 7a-l, computational methods and crystallographic data of (*S*)-7a and (*R*)-7e. This material is available free of charge via the Internet at http:// pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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